#### USAF School of Aerospace Medicine & Defense Health Agency

# DoD Global, Laboratory-Based, Influenza Surveillance Program

## 2016-2017















#### **Cumulative Results**

Locations	77
Collected	2,266
Tested	2,122

	_
Influenza A	541
A(H1N1)pdm09	4
A(H3N2)	536
A(H3N2) & RSV	1
A/not subtyped	0
	•
Influenza B*	40
В	40

# Respiratory Highlights 22 January - 4 February 2017 (Surveillance Weeks 4 & 5)

- During 22 January 4 February 2017, a total of 460 specimens were collected from 52 locations. Results were finalized for 357 specimens from 51 locations. During Week 4, two influenza A(H1N1)pdm09, 92 influenza A(H3N2), and ten influenza B viruses were identified. During Week 5, 75 influenza A(H3N2) and ten influenza B viruses were identified. Approximately 40% of specimens tested positive for influenza during Week 4. Approximately 43% of specimens tested positive for influenza during Week 5. The influenza percent positive for the season is approximately 27%.
- According to the CDC, influenza activity has continued to increase in the United States.
   Twenty influenza-associated pediatric deaths have been reported for the 2016-2017 season.
   The proportion of people who sought medical care for influenza-like illness (ILI) has been at or above the national baseline of 2.2% for eight consecutive weeks (CDC, FluView Report Week 5, cited 10 February 2017).

Other Respiratory Pathogens	627
Adenovirus	38
Bordetella pertussis	0
Chlamydophila pneumoniae	2
Coronavirus	62
Human Metapneumovirus	27
Mycoplasma pneumoniae	27
Parainfluenza	113
RSV	108
Rhino/Enterovirus	169
Non-influenza Viral Coinfections	73
Non-influenza Bacterial Coinfections	8
-M. pneumo coinfections (8)	

Lab data are current as of 6 February 2017. Results are preliminary and may change as more results are received.

\*Influenza B lineages will be reported in the periodic molecular sequencing reports.

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Table 1. Results by region and location for specimens collected during Weeks 4 & 5

Region*		A(H1N1)pdm09	A(H3N2)	В	Adenovirus	Coronavirus	NNMH	Parainfluenza	RSV	Rhinovirus/Enterovirus	Adeno & Corona & RSV	Adeno & Rhino/Entero	Corona & RSV	Corona & RSV & Rhino/Entero	hMNV & Rhino/Entero	Para & RSV	RSV & Rhino/Entero	No Pathogen	Total
Deployed	Country 1, Location B	-	-	-	-	ı	-	-	-	-	ı	-	-	-	-	-	-	1	1
	Country 2, Location A	-	4	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	5
EUCOM	Incirlik AB, Turkey	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	1	2
PACOM	Eielson AFB, AK	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	1
	Kadena AB, Japan	-	-	-	-	1	-	-	-	-	1	-	-	-	-	-	-	1	1
	Yokota AB, Japan	-	4	-	-	1	1	-	-	-	1	-	-	-	-	-	-	1	6
Region 1	Hanscom AFB, MA	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	2
	USCG Academy, CT	-	1	-	-	-	-	-	_	-	-	-	-	-	-	-	-	2	3
Region 2	Ft Drum, NY	-	-	5	-	-	1	-	-	-	-	-	-	-	-	-	-	-	6
	JB M cGuire-Dix-Lakehurst, NJ	-	9	-	-	-	2	-	-	2	-	-	-	-	-	-	1	8	22
	USM A - West Point, NY	-	12	-	-	2	-	1	1	-	-	-	-	-	-	-	-	10	26
Region 3	Dover AFB, DE	-	2	-	-	-	-	-	_	-	-	-	-	-	-	-	-	5	7
	JB Anacostia-Bolling, DC	-	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2
	JB Andrews, M D	-	6	1	-	-	-	-	_	-	-	-	-	-	-	-	-	4	11
	JB Langley-Eustis, VA	_	7	_	_	1	-	-	4	-	_	_	_	-	-	_	_	10	22
	US Naval Academy, M D	_	1	-	_	-	-	-	_	_	_	_	_	-	-	_	_	-	1
Region 4	Columbus AFB, MS	_	1	-	_	1	-	-	_	_	-	_	_	_	_	_	_	_	2
	Eglin AFB, FL	_	3	-	_	-	-	-	_	_	_	_	_	-	-	_	_	_	3
	Ft Bragg, NC	-	1	_	-	_	-	_	_	-	_	_	-	-	_	-	-	7	8
	Ft Campbell, KY	1	6	1	1	_	-	-	_	_	_	-	_	_	1	_	_	4	14
	Hurlburt Field, FL	Ė	2	Ė	-	-	-	-	_	-	-	-	_	_	<u>-</u>	_	-	1	3
	Keesler AFB, MS	-	-	-	-	-	-	-	_	1	-	-	_	_	-	_	-	<u> </u>	1
	M axwell AFB, AL	-	<b>-</b>	<u> </u>	_	_	<u> </u>	<b>-</b>	-	Ė	_	<u> </u>	<u> </u>	<del>  -</del>	<u> </u>	<u> </u>	l _	1	1
	M oody AFB, GA	-	5	<u> </u>	_	_	l	<b>-</b>	-	2	_	<u> </u>	<u> </u>	<b> </b>	<u> </u>	<u> </u>	<b>-</b>	Ė	7
	NH Jacksonville, FL	-	1	<del>  -</del>	_	_	-	-	_	<u>-</u>	_	_	_	_	  -	_	_	_	1
	Robins AFB, GA	-	5	2	_	-	-	<u> </u>	-	_		<u> </u>	-		<u> </u>	-	_	3	10
	Seymour Johnson AFB, NC		2	╘			-	Ė										<u> </u>	2
	Shaw AFB, SC		4	2	Ē	Ē	_	Ė	<u> </u>	Ė	Ē	Ē	<u> </u>	<u>-</u>	Ē	<u> </u>	Ē	1	7
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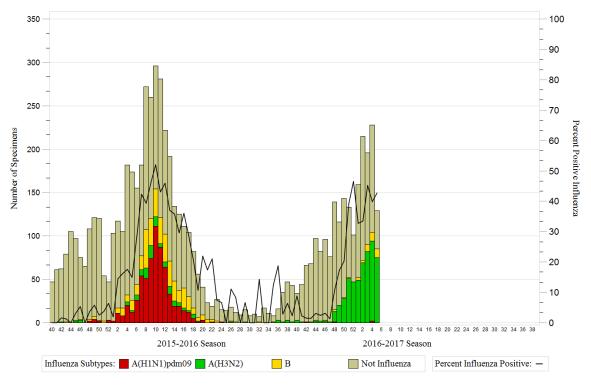
(Cont'd from page 3)

Table 1. Results by region and location for specimens collected during Weeks 4 & 5 (Cont'd from page 2)

Region*		A(H1N1)pdm09	A(H3N2)	В	Adenovirus	Coronavirus	NM NV	Parainfluenza	RSV	Rhinovirus/Enterovirus	Adeno & Corona & RSV	Adeno & Rhino/Entero	Corona & RSV	Corona & RSV & Rhino/Entero	hMNV & Rhino/Entero	Para & RSV	RSV & Rhino/Entero	No Pathogen	Total
Region 5	Wright-Patterson AFB, OH	-	_	_	-	-	-	_	-	-	-	_	-	-	_	1	-	1	2
Region 6	Altus AFB, OK	-	4	-	-	-	-	-	2	-	1	-	-	-	-	-	1	3	11
	Barksdale AFB, LA	-	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-	1
	Cannon AFB, NM	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	3	3
	Ft Polk, LA	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1
	Little Rock AFB, AR	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2	2
	Sheppard AFB, TX	-	20	-	-	-	-	1	-	1	-	-	-	-	-	-	-	5	27
	Tinker AFB, OK	-	14	1	-	1	-	-	-	-	-	-	-	-	-	-	-	6	22
	Vance AFB, OK	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	1
Region 7	M cConnell AFB, KS	-	3	-	-	2	-	-	1	-	-	-	-	-	-	-	-	1	7
	Offutt AFB, NE	1	11	2	1	1	1	-	-	1	1	-	-	-	-	1	-	2	18
Region 8	Ellsworth AFB, SD	-	3	-	1	1	1	-	-	-	1	-	-	-	-	-	1	2	5
	FE Warren AFB, WY	-	8	1	1	2	1	1	2	1	1	-	-	-	-	-	-	3	18
	Hill AFB, UT	-	1	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	2
	M almstrom AFB, M T	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1
	M inot AFB, ND	-	5	-	-	-	-	-	1	-	-	1	-	-	-	-	-	1	8
	Peterson AFB, CO	-	4	1	-	1	-	-	3	-	-	-	-	-	-	-	-	1	10
Region 9	Beale AFB, CA	-	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2
	Davis-M onthan AFB, AZ	-	1	-	-	-	-	-	-	1	-	-	1	1	-	-	1	-	5
	Nellis AFB, NV	-	-	1	-	-	-	-	-	1	-	-	-	-	-	-	-	2	4
	Travis AFB, CA	-	5	-	-	-	1	-	3	-	-	-	-	-	-	-	-	2	11
Region 10	Fairchild AFB, WA	-	2	-	1	-	-	-	-	-	-	-	-	-	-	-	-	2	5
	Mt Home AFB, ID	-	4	-	-	-	-	3	2	-	-	-	2	_	-	-	-	3	14
Total		2	167	20	2	11	5	7	20	10	1	1	3	1	1	1	3	102	357

#### **Laboratory Results - Cumulative for Season**

**Graph 1.** Percent influenza positive by week: 2015-2016 surveillance year and through Week 5 of the 2016-2017 surveillance year



Note: Dual influenza coinfections are excluded from this graph. Specimens with pending results are used in the denominator to calculate percent positive, but are not displayed in the graph.

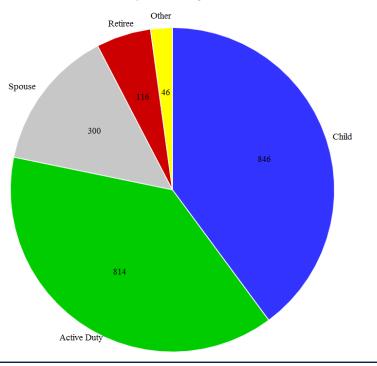
**Table 2.** ILI by age group for the 2016-2017 surveillance year through Week 5

Age Group	Frequency	Percent
0-5	488	23.00
6-9	136	6.41
10-17	223	10.51
18-24	331	15.60
25-44	699	32.94
45-64	189	8.91
65+	56	2.64

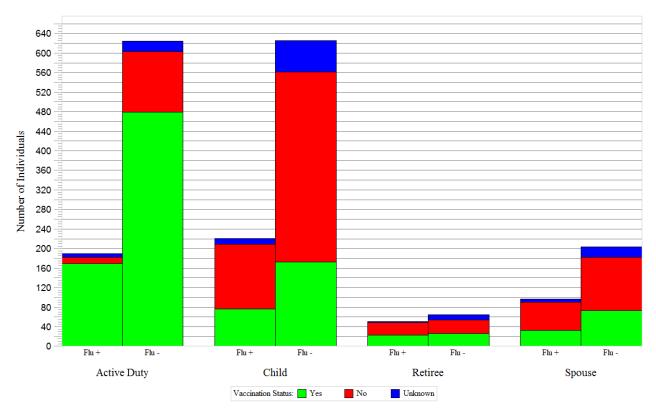
#### **Demographic Summary**

Of 2,122 ILI cases, 814 (38.4%) are service members, 846 (39.9%) are children, 300 (14.1%) are spouses, and 162 (7.6%) are retirees and other beneficiaries. The median age of ILI cases with known age (n=2,122) is 22 (range 0, 96).

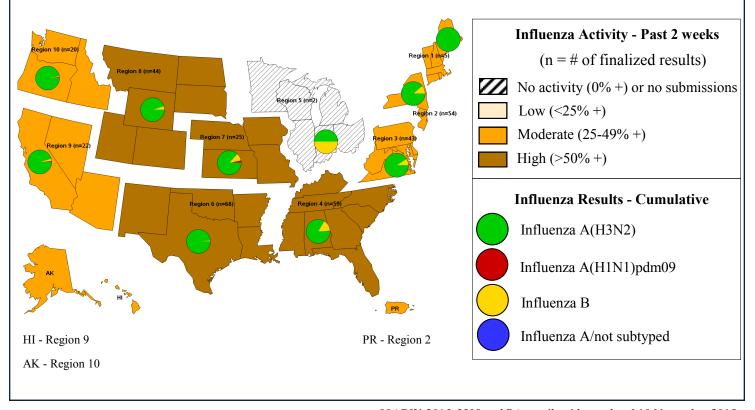
**Graph 2.** ILI by beneficiary status for the 2016-2017 surveillance year through Week 5



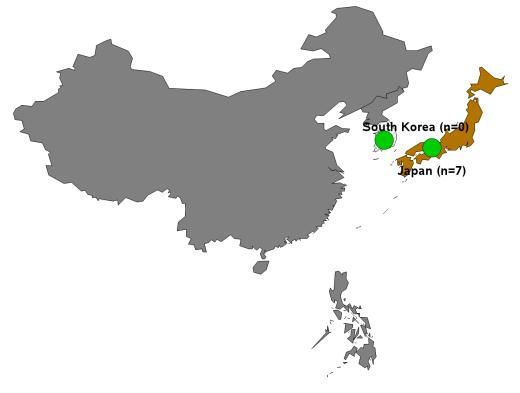
**Graph 3.** Vaccination status by beneficiary type for the 2016-2017 surveillance year through Week 5



Map 1. Influenza subtypes and activity level by U.S. region for the 2016-2017 surveillance year through Week 5

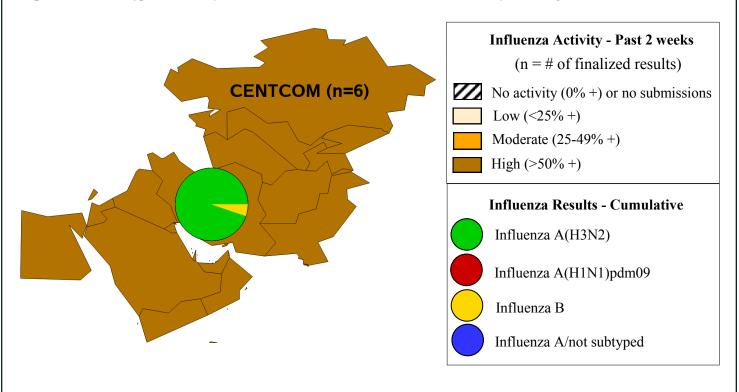


Map 2. Influenza subtypes and activity level by country for the 2016-2017 surveillance year through Week 5 (Pacific)



Note - Countries shaded in gray do not contain sentinel sites and are only displayed for geographical perspective.

Map 3. Influenza subtypes and activity level for CENTCOM for the 2016-2017 surveillance year through Week 5



Note - Specimens for CENTCOM were tested at USAFSAM or Landstuhl Regional Medical Center (LRMC).

## Laboratory Results—Through Current Surveillance Week 5

Table 3. Cumulative results by region and location for specimens collected during the 2016-2017 surveillance year

Region*		A(H1N1)pdm09	A(H3N2)	A(H3N2) & RSV	В	Adenovirus	C. pneumoniae	Coronavirus	NMMV	M. pneumoniae	Parainfluenza	RSV	Rhinovirus/Enterovirus	Non-Influenza Viral Coinfection	Non-Influenza Bacterial Coinfection	No Pathogen	Total
Deployed	Country 1, Location A	-	3	-	-	-	-	1	-	1	-	1	-	-	-	7	12
	Country 1, Location B	-	6	-	1	-	-	1	-	-	-	1	-	1	-	5	15
	Country 1, Location D	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	1
	Country 2, Location A	-	27	-	1	-	-	5	-	-	-	-	4	2	-	10	49
EUCOM	Incirlik AB, Turkey	-	1	-	1	-	-	-	-	-	-	-	-	-	-	1	3
PACOM	CFA Okinawa, Japan	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2	2
	Eielson AFB, AK	-	-	-	-	-	-	-	-	1	-	-	-	-	-	5	6
	JB Elmendorf-Richardson, AK	-	1	-	-	-	-	-	-	-	-	-	1	-	-	1	3
	JR M arianas - Andersen AFB, Guam	-	-	-	-	-	-	-	-	1	-	-	-	-	-	1	2
	Kadena AB, Japan	-	3	-	-	-	-	-	-	-	2	-	2	-	1	18	26
	Kunsan AB, South Korea	-	2	-	-	-	-	1	-	-	-	-	1	-	-	1	5
	M isawa AB, Japan	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	1
	Osan AB, South Korea	-	3	-	-	-	-	-	-	-	-	-	-	-	-	6	9
	Tripler AM C, HI	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	1
	Yokota AB, Japan	-	14	-	-	1	-	2	1	2	-	2	7	3	-	42	74
Region 1	Hanscom AFB, MA	-	1	-	-	-	-	-	-	1	1	-	-	1	-	2	6
	USCG Academy, CT	-	4	-	-	-	-	-	-	1	-	-	3	-	2	4	14
Region 2	CGAS Borinquen, PR	-	1	_	-	_	-	-	-	-	-	_	-	-	-	-	1
	Ft Drum, NY	-	1	_	8	4	-	1	7	1	6	4	1	2	-	10	45
	JB M cGuire-Dix-Lakehurst, NJ	-	30	-	-	2	-	5	3	2	7	5	9	4	-	51	118
	USM A - West Point, NY	Ι.	40	_	1	10	_	4	2	1	5	9	4	4	_	80	160
Region 3	Dover AFB, DE	-	5	_	Ė	1	-	1	-	1	-	1	1	-	-	15	25
<b>g</b>	JB Anacostia-Bolling, DC	-	2	_	_	-	_	Ė	-	Ė	_	1	-	_	_	-	3
	JB Andrews, M D		10	_	1	_		1			_	2	1	2		19	36
	JB Langley-Eustis, VA	-	12	-	2	-	-	2	2	2	4	17	22	8	_	57	128
	NM C Portsmouth, VA	H	-	-		-	_				_	17	1	-	_	2	3
	US Naval Academy, M D	Ė	1	-	-	-	_	-	Ė	Ε.	-	-	-	-	_	-	1
Region 4	Columbus AFB, M S	-	3	_	-	-	-	1	-	-	-	_	1	-	_	7	12
. tog.o	Eglin AFB, FL	H	8	-	1	2	-	1	-	-	-	4	9	3	_	16	44
	Ft Bragg, NC	-	4	-	3	-	-	1	-	1	3	2	6	3	3	21	47
	Ft Campbell, KY	۲.				1		H	1	_	,	3	-	5	-	10	37
	Hurlburt Field, FL	1	7	-	2	-	-	H	_	-	1	3	-	-	-	7	15
	JB Charleston (AF), SC	-	3	-	-	-	-	-	-	-	-	-	-	-	-	2	5
	Keesler AFB, MS	H	3	-	-	-	-	H	-	-	1	2	1	1	-	7	
	M acDill AFB, FL	H	-	-	-	-	-	-	Ė	÷	-			-	-		12
	M axwell AFB, AL	ŀ	2	Ė	Ė	Ë	Ė	Ė	Ė	Ė	1	Ė	1	Ė	Ė	8	12
	M oody AFB, GA	-	11	Ė	2	1	-	1	-	-	1	5	4	7	1	16	49
	NH Beaufort, SC	Ė	- 11	Ė	_	<u> </u>	Ė	H	Ė	Ė	<del> </del>	٥	+		_		
	NH Camp Lejeune, NC	H	1	H	Ė	Ε-	Ė	H	H	H	H	Η-	Ε-	Ė	Ė	2	2
	NH Jacksonville, FL	H		H	-	H	-	H	-	-	H	<u> </u>	H	-	-	4	5
	Patrick AFB, FL	Ė	1	<del>-</del>	<u> </u>	-	Ė	H	i –	H	H	-	-	Ė	Ė	2	3
	Robins AFB, GA	-	-	-	-	-	-	-	<u> </u>	_	-	-	-	-	-	1	1
	,	-	10	-	2	-	-	-	-	1	-	1	-	-	-	7	21
	Seymour Johnson AFB, NC	-	2	-	-	1	-	-	-	1	1	-	1	-	-	3	9
	Shaw AFB, SC	-	11	-	4	1	-	5	1	1	3	1	3	-	-	27	57

(Cont'd on page 8)

\*CONUS locations are based on Health & Human Services regions. Other locations are defined by COCOM.

## Laboratory Results—Through Current Surveillance Week 5

Table 3. Cumulative results by region and location for specimens collected during the 2016-2017 surveillance year (Cont'd from page 7)

Region*		A(H1N1)pdm09	A(H3N2)	A(H3N2) & RSV	В	Adenovirus	C. pneumoniae	Cor onavir us	hMNV	M. pneumoniae	Parainfluenza	RSV	Rhinovirus/Enterovirus	Non-Influenza Viral Coinfection	Non-Influenza Bacterial Coinfection	No Pathogen	Total
Region 5	Scott AFB, IL	-	-	-	1	-	-	-	-	1	2	1	1	-	1	6	13
	Wright-Patterson AFB, OH	-	1	-	-	-	-	-	-	1	1	-	1	2	-	10	16
Region 6	Altus AFB, OK	-	4	-	-	1	-	-	1	-	-	3	4	2	-	22	37
	Barksdale AFB, LA	-	-	-	-	-	-	-	-	-	2	-	1	-	-	4	7
	Cannon AFB, NM	-	2	-	-	-	-	1	-	1	1	-	3	-	-	14	22
	Ft Polk, LA	-	1	-	-	-	-	-	-	1	-	-	-	-	-	1	3
	Laughlin AFB, TX	-	-	-	-	-	-	-	2	-	-	-	-	-	-	1	3
	Little Rock AFB, AR	-	1	-	-	-	-	-	-	-	-	-	-	-	-	2	3
	Sheppard AFB, TX	-	34	-	-	-	-	3	1	-	6	-	6	-	-	30	80
	Tinker AFB, OK	-	33	1	1	-	-	2	-	1	5	4	6	-	-	40	93
	Vance AFB, OK	-	-	-	-	-	-	-	-	-	-	-	-	-	-	10	10
Region 7	M cConnell AFB, KS	-	10	-	-	-	-	3	-	1	2	1	4	-	-	15	36
	Offutt AFB, NE	1	16	-	3	1	-	3	-	-	-	-	5	1	-	24	54
Region 8	Ellsworth AFB, SD	-	7	-	-	-	-	2	-	-	3	-	3	-	-	14	29
	FE Warren AFB, WY	-	16	-	1	2	-	2	-	1	2	4	3	-	-	16	47
	Hill AFB, UT	-	17	-	-	-	-	2	-	-	4	3	4	-	-	22	52
	M almstrom AFB, M T	-	4	-	1	-	-	-	-	1	-	-	1	-	-	3	10
	M inot AFB, ND	-	5	-	-	-	-	1	1	1	-	1	2	1	-	6	18
	Peterson AFB, CO	-	9	-	1	-	-	2	-	-	1	6	3	4	-	10	36
	USAF Academy, CO	-	1	-	-	-	-	-	1	-	-	1	1	-	-	2	6
Region 9	Beale AFB, CA	-	5	-	-	-	-	-	-	-	-	-	-	-	-	-	5
	Davis-M onthan AFB, AZ	-	6	-	-	-	-	-	-	-	5	-	4	3	-	14	32
	Edwards AFB, CA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2	2
	Luke AFB, AZ	-	-	-	-	-	-	2	-	-	-	1	-	1	-	4	8
	Nellis AFB, NV	1	2	-	2	2	-	1	-	-	4	3	5	6	-	15	41
	Travis AFB, CA	-	49	-	-	-	1	1	4	-	6	5	8	1	-	29	104
	Vandenberg AFB, CA	-	-	-	-	-	-	-	-	-	-	-	-	-	-	3	3
Region 10	CGS North Bend, OR	_	2	-	-	-	-	-	-	-	-	-	-	-	-	2	4
	Fairchild AFB, WA	1	13	-	1	2	-	3	-	-	4	2	4	-	-	38	68
	JB Lewis-M cChord, WA	-	2	-	-	1	-	-	-	-	-	-	-	-	-	-	3
	Mt Home AFB, ID	-	19	-	-	1	1	1	-	1	23	8	12	4	-	56	126
	NH Bremerton, WA	-	33	-	-	4	-	-	-	-	6	4	4	2	-	14	67
Total		4	536	1	40	38	2	62	27	27	113	108	169	73	8	914	2122

\*CONUS locations are based on Health & Human Services regions. Other locations are defined by COCOM.

#### **Molecular Sequence Analysis Report #3**

## **USAFSAM Epidemiology Laboratory Service**

This is the third USAFSAM influenza sequence surveillance report for the 2016-2017 influenza season and includes a total of 75 specimens collected between 12 July 2016 and 9 January 2017, with 66 specimens analyzed by USAFSAM, six hemagglutinin sequences provided by the Naval Medical Research Unit 6 (NAMRU-6) in Peru, and three hemagglutinin sequences provided by the Walter Reed Army Institute of Research (WRAIR). Among the specimens analyzed, two (2.7%) were influenza A(H1N1)pdm09, 65 (86.7%) were influenza A(H3N2), four (5.3%) were influenza B/Victoria lineage, and four (5.3%) were influenza B/Yamagata lineage. Figure 1 shows the proportion of hemagglutinin (HA) sequences analyzed for this report by type/subtype, USMAJCOM, and influenza A(H3N2)-specific clade designations. Sequences for each type and subtype included in this report are shown for each sentinel site in Table 1. Figures 2-5 display the phylogenetic relationships among HA sequences for influenza A(H1N1)pdm09, A(H3N2), and influenza B/Victoria and B/Yamagata lineages, respectively.

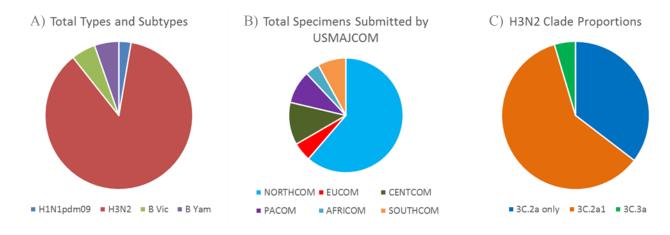


Figure 1: A) Proportion of influenza hemagglutinin sequence subtypes and lineages submitted to USAFSAM for this report, B) Number of influenza positive specimens submitted by sentinel sites separated by USMAJCOM (AFRICOM data from WRAIR, SOUTHCOM data from NAMRU-6), C) Proportion of specimens in each influenza A(H3N2) clade represented in this report.

\*CONUS locations are based on Health & Human Services regions. Other locations are defined by COCOM.

**Table 1**: Influenza subtypes and lineages from corresponding sentinel sites included in the analyses for this report.

	A(H1N1)pdm09	A(H3N2)	B /Victoria	B /Yamagata	Grand Total
CONUS					
California					
Travis AFB		9			9
Colorado					
Peterson AFB		1			1
Florida					
Eglin AFB		1			1
Georgia					
Moody AFB		1			1
Robins AFB		1			1
Idaho					
Mt Home AFB		2			2
Kansas					
McConnell AFB		1			1
Maryland					
JB Andrews		1			1
Nebraska					
Offutt AFB		1			1
New York					
USMA - West Point		2			2
North Carolina					
Ft Bragg		2			2
Ohio					
Wright-Patterson AFB	1	1	2	1	5
Oklahoma					
Tinker AFB		3			3
South Carolina					
Shaw AFB		1			1
Texas					
SAMMC		4	1		5
Sheppard AFB		2			2
Utah					
Hill AFB		4			4
Virginia					
JB Langly-Eustis				1	1
Washington					
Fairchild AFB		2			2
JB Lewis-McChord		1			1

(Cont'd on page 11)

**Table 1**: Influenza subtypes and lineages from corresponding sentinel sites included in the analyses for this report. *(Cont'd from page 10)* 

	A(H1N1)pdm09	A(H3N2)	B/Victoria	B/Yamagata	Grand Total
OCONUS					
Country1					
Location A		2			2
Country2					
Location A		7			7
Germany					
Landstuhl RMC		2		1	3
Vilseck AHC		1			1
Japan					
Yokota AB		2			2
Nigeria					
WRAIR		3			3
Paraguay					
NAMRU-6	1		1		2
Peru					
NAMRU-6		3		1	4
South Korea					
Brian Allgood ACH		4			4
Kunsan AB		1			1
Grand Total	2	65	4	4	75

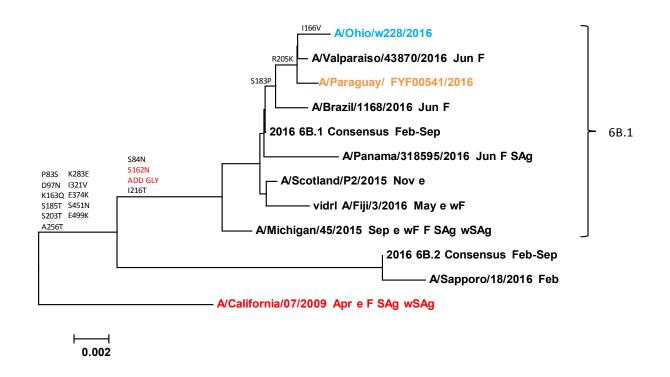
The hemagglutinin (HA) gene from select influenza positives was sequenced using dye terminator, Sanger-based methods. Preliminary data are based on the sequence analysis of the hemagglutinin gene. Antigenic sites, receptor binding sites and glycosylation motifs are predicated upon correlations with previously published experimental evidence. Sequence data was constructed and analyzed using multiple software programs. Genetic and predicted antigenic information that resulted from this analysis is shared with United States Centers for Disease Control and Prevention (CDC), World Health Organization (WHO) and contribute to the seasonal Northern and Southern Hemisphere vaccine component selections.

### Influenza A(H1N1)pdm09

- Among the 67 influenza A isolates, two (3.0%) were influenza A(H1N1)pdm09. The influenza A(H1N1)pdm09 sequences are characterized in a neighbor-joining phylogenetic tree with reference strains rooted from the current vaccine strain, A/California/07/2009-like virus (Figure 2).
- The A(H1N1)pdm09 isolates characterized for this report exhibited an overall protein homology of 97.2%-97.3% compared to the 2016-2017 influenza vaccine component, A/California/07/2009-like virus.
- Both of the A(H1N1)pdm09 HA sequences for this report contain mutations consistent with the predominating subgroup, referred as group 6B, and all classified as clade 6B.1 (distinguished by the mutations S162N and I216T).
- Gain or loss of *N*-linked glycosylation sites has been shown to alter HA protein surface topology. A gain in glycosylation could be advantageous to the virus by virtue of a masking effect on important antibody recognition sites, thus potentially modulating viral antigenicity. Observations are based solely on sequence motifs. For the influenza A(H1N1)pdm09 isolates characterized in this report, one mutation, S162N (serine to asparagine), was observed that could cause a gain of a glycosylation motif.
- Of the 17 mutations present in the A(H1N1)pdm09 isolates, five occurred at predicted antigenic sites (zero at site A, one at site B, zero at site C, two at site D, and two at site E) and two occurred at the receptor binding site.<sup>2,5</sup>

Figure 2. Influenza A(H1N1)pdm09 HA Phylogenetic Analysis

# Current 2016-2017 A(H1N1)pdm09 Vaccine strain: A/California/07/2009 Reference Strain August 2016 December 2016 ADD GLY Create Glycosylation Motif F - CDC Reference Antigen WF - WHOcc Reference Antigen SAg - Serology Antigen wSAg - WHOcc Serology Antigen e Egg Isolate



#### Influenza A(H3N2)

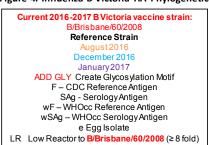
- Among the 67 influenza A isolates, 65 (97.0%) were influenza A(H3N2). The influenza A(H3N2) HA sequences are characterized in a neighbor-joining phylogenetic tree with reference strains rooted from a previous vaccine strain, A/Texas/50/2012 (Figure 3).
- The A(H3N2) isolates characterized for this report exhibited an overall protein homology of 97.3%-99.1% (average 98.2%) compared to the 2016-2017 influenza vaccine component, A/Hong Kong/4801/2014-like virus.
- All of the influenza A(H3N2) isolates sequenced for this report were in clade 3C. Sixty-two (95.4%) of the influenza A(H3N2) sequences classified as subclade 3C.2a and 3 (4.6%) classified as subclade 3C.3a. Thirty-nine of the 62 3C.2a isolates (62.9%) further classified as the newly distinguished subclade within 3C.2a, 3C.2a1 (determined by the mutations N171K, I406V, and G484E). Another mutation of interest, N121K, was present in 28 (71.8%) of the 3C.2a1 isolates (43.1% of the total H3N2).
- Among the influenza A(H3N2) isolates characterized in this report, seven mutations: N122K (asparagine to lysine), N122D (asparagine to aspartic acid), T128N (threonine to asparagine), T135K (threonine to lysine), N144S (asparagine to serine), T160I (threonine to isoleucine), and T160K (threonine to lysine) were observed that could cause the loss of a glycosylation motif. Two other mutations, N128T (asparagine to threonine) and K160T (lysine to threonine), were observed that could cause the gain of a glycosylation motif.
- Of the 59 mutations present in the A(H3N2) specimens, 21 occurred at predicted antigenic sites (six at site A, four at site B, three at site C, three at site D, and five at site E) and two occurred at the receptor binding site.<sup>2,5</sup>
- A/California/1402/2017 was collected from a hospitalized patient who was not vaccinated. Records show that the patient exhibited flu-like symptoms which included fever (102.5°F), cough, fatigue, body aches, chills, runny nose, and headache at the time this specimen was collected.

#### Global, Laboratory-Based, Influenza Surveillance Program S46T A/California/1209/2016 A/NewYork/1201/2016 M168I A/Germany/x76/2016 Figure 3. Influenza A(H3N2) HA Phylogenetic Analysis A/Washington/1353/2017 N96S A/California/1376/2017 A106T G142R N122K LOSS GLY A/Washington/1107/2016 Current 2016-2017 A(H3N2) Vaccine strain: A/Germany/x9/2016 A/Hong Kong/4801/2014 A/Oklahoma/1049/2016 A/Oklahoma/1251/2016 T135K LOSS GLY Reference Strain G479E I A/SouthKorea/x53/2016 A/SouthKorea/x54/2016 August 2016 A/SouthKorea/x56/2016 T160 October 2016 A/Oklahoma/1415/2017 November 2016 A/Washington/1451/2017 December 2016 148K 1260M A/SouthKorea/1159/2016 A/Japan/1177/2016 A/Country1/1141/2016 January 2017 ADD GLY Create Glycosylation Motif N121K A/Country1/747/2016 A/Ohio/w187/2016 A/Ohio/w187/2016 A/California/1208/2016 LOSS GLY Loss of Glycosylation Motif F – CDC Reference Antigen D53F wF - WHOcc Reference Antigen A/Texas/1393/2017 A/SouthKorea/x51/20 SAg - Serology Antigen N3789 3C.2a1 vidrl A/Newcastle/30/2016 Mar wSAg wSAg - WHOcc Serology Antigen A/Texas/x48/2016 F193S MNAg - Microneutralization Antigen K92R H3110 A/Kansas/1196/2016 FRA – Focus Reduction Antigen A/NewYork/1436/2017 0197R A/Texas/1463/2017 e Egg Isolate HKLR - Low Reactor to 2016 HA 3C2a Consensus Feb-Sep A/Hong Kong/4801/2014-like Cell (≥ 8 fold) A/California/1400/2017 T160I LOSS GLY A/Utah/1128/2016 K2R Q80K G142R A/California/1202/2016 N31S A/Country2/1094/2016 1214T A/Texas/x43/2016 R545K K142F A/Colorado/1186/2016 K92R 3C.2a niid A/Nagano-C/21/2016 Feb wSAg N171K A/Alaska/232/2015 Sep F MNAg wSAg 1406V A/California/1369/2017 V529F A/Utah/1397/2017 cnic A/Heilongjiang-Longsha/1274/2016 Apr wSAg R142K crick A/Hong Kong/97/2016 Jan wSAg A/Maryland/1429/2017 A212T A212T A/Florida/1234/2017 T131K A/Nebraska/124 A/Georgia/1384/2017 - A/Utah/1164/2016 Q197R A/Georgia/1390/2017 A/SouthCarolina/1192/2016 A/Idaho/1124/2016 D188N A/NorthCarolina/1324/2017 A/NorthCarolina/1326/2017 A/New Jersey/26/2014 Nov NC3 F A/Country2/1312/2017 N121K A/Country2/1314/2017 T160K LOSS GLY S144K A/Nigeria/1514A03006826T/2016 K450R| A/Nigeria/1514A03007000T/2016 A/Nigeria/1514A03006993T/2016 A/Germany/x78/2016 G275S K160T A/Country2/1216/2016 N128T ADD GLY A/Country2/1327/2017 N144S LOSS GLY A/Michigan/15/2014 Sep F SAg F150V crick A/South Africa/R2651/2015 May wF Q311H A/Peru/FPI15434/2016 T128N LOSS GLY Peru/PBA27210148/2016 D489N A/Peru/FPP02924/2016 P198S N128A N225D A138S A/North Dakota/19/2016 Mar FRA MNAg R142G F159S A/Hong Kong/4801/2014 Feb e wF F SAg MNAg wS A/Switzerland/9715293/2013 Dec e F wF SAg wSAg K326R A/Texas/88/2016 Feb FRA HKLR F SAg MNAg wSAg S198P S312N 2016 HA 3C3a Consensus Feb-Sep A/California/1375/2017 3C.3a A/Sydney/142/2016 Jun A/California/1402/2017 \* HOSPITALIZED A/Texas/50/2012 F 0.002

#### Influenza B

- The influenza B isolates are characterized in lineage specific, neighbor-joining phylogenetic trees with reference strains and are midpoint rooted for both the B/Victoria isolates (Figure 4) and the B/Yamagata isolates (Figure 5).
- The distinguishing characteristic between the two influenza B lineages (Victoria & Yamagata) is defined by an amino acid deletion in viruses belonging to the Yamagata lineage. Four (50.0%) of the influenza B isolates characterized in this report fell into the Victoria lineage and four (50.0%) fell into the Yamagata lineage.
- The influenza B/Victoria isolates characterized for this report exhibited a protein homology of 98.7% when compared to the 2016-2017 B/Victoria vaccine component, B/Brisbane/60/2008-like virus. The influenza B/Yamagata isolates characterized for this report exhibited a protein homology of 98.9%-99.1% (average 90.0%) when compared to the 2016-2017 B/Yamagata vaccine component, B/Phuket/3073/2013-like virus.
- All of the influenza B/Victoria isolates fall into clade V1A and all of the B/Yamagata isolates fall into clade Y3
- For B/Victoria, one mutation, A199T (alanine to threonine), adds a glycosylation motif. For B/Yamagata, one mutation, D197N (aspartic acid to asparagine), adds a glycosylation motif.

Figure 4. Influenza B Victoria HA Phylogenetic Analysis



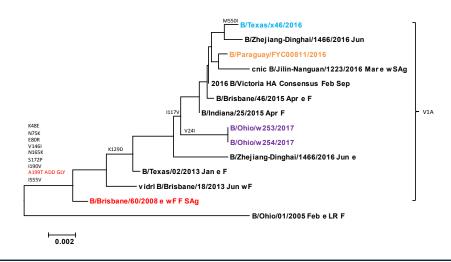
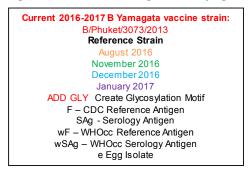
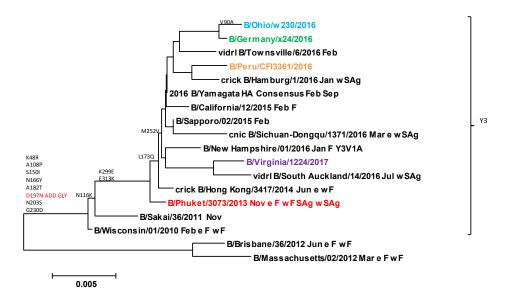


Figure 5. Influenza B Yamagata HA Phylogenetic Analysis





#### **References:**

- 1. Wright, P., Neumann, G., and Kaqaoka, Y. (2007). Orthomyxoviruses In: Knipe, D.M., Howley, P.M. (Eds.), Fields Virology. Wolters Kluwer, Lippincott Williams & Wilkins, Philadelphia, pp.1692-1740.
- 2. Kongchanagul, A., Suptawiwat, O., Kanrai, P., Uiprasertkul, M., Puthavathana, P., and Auewarakul P. (2008). Positive selection at the receptor-binding site of hemagglutinin H5 in viral sequences derived from human tissues. Journal of Gen. Vir. **89**, 1805-1810.
- 3. Cherry, J.L., Lipman, D.J., Nikolskaya, A., and Wolf, Y.I. (2009). Evolutionary Dynamics of N-Glycosylation Sites of Influenza Virus Hemagglutinin. *PLoS Curr Influenza*. August 18: RRN1001.
- 4. Deem, M., and Pan, K. (2009). The epitope regions of H1-subtype influenza A, with application to vaccine efficacy. Protein Engineering, Design and Selection. **22**, no. 9. 543-546.
- 5. Wolf, Y.I., Viboud, C., Holmes, E.C., Koonin, E.V., and Lipman, D.J. (2006). Long intervals of stasis punctuated by bursts of positive selection in the seasonal evolution of influenza A virus. *Biol Direct*.; 1: 34. doi: 10.1186/1745-6150-1-34.

#### **References:**

- 1. Wright, P., Neumann, G., and Kaqaoka, Y. (2007). Orthomyxoviruses In: Knipe, D.M., Howley, P.M. (Eds.), Fields Virology. Wolters Kluwer, Lippincott Williams & Wilkins, Philadelphia, pp.1692-1740.
- 2. Kongchanagul, A., Suptawiwat, O., Kanrai, P., Uiprasertkul, M., Puthavathana, P., and Auewarakul P. (2008). Positive selection at the receptor-binding site of hemagglutinin H5 in viral sequences derived from human tissues. Journal of Gen. Vir. **89**, 1805-1810.
- 3. Cherry, J.L., Lipman, D.J., Nikolskaya, A., and Wolf, Y.I. (2009). Evolutionary Dynamics of N-Glycosylation Sites of Influenza Virus Hemagglutinin. *PLoS Curr Influenza*. August 18: RRN1001.
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#### **Background**

The DoD-wide program was established by the Global Emerging Infections Surveillance and Response System (GEIS) in 1997. The surveillance network includes the Defense Health Agency/Armed Forces Health Surveillance Branch—Air Force Satellite Cell (DHA/AFHSB-AF) and U.S. Air Force School of Aerospace Medicine (USAFSAM) (sentinel site respiratory surveillance), the Naval Health Research Center (recruit and shipboard population-based respiratory surveillance), the Naval Medical Research Unit (NAMRU-3) in Cairo, Egypt, the Naval Medical Research Unit (NAMRU-2) in Phnom Penh, Cambodia, the Armed Forces Research Institute of Medical Sciences (AFRIMS) in Bangkok, Thailand, the Naval Medical Research Unit (NAMRU-6) in Lima, Peru, and the United States Army Medical Research Unit-Kenya (USAMRU-K) located in Nairobi, Kenya. This work is supported by the Air Force and GEIS Operations, a Division of the Armed Forces Health Surveillance Branch (AFHSB).

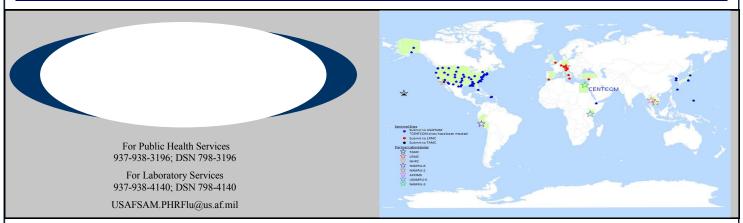
#### **Sentinel Site Surveillance**

In 1976, the U.S. Air Force Medical Service began conducting routine, global, laboratory-based influenza surveillance. Air Force efforts expanded to DoD-wide in 1997. DHA/AFHSB-AF and USAFSAM manages the surveillance program that includes global surveillance among DoD beneficiaries at over 95 sentinel sites (including deployed locations) and many non-sentinel sites (please see map below). Collaborating partner laboratories include five DoD overseas medical research laboratories (AFRIMS, NAMRU-2, NAMRU-3, NAMRU-6, USAMRU-K) who collect specimens from local residents in surrounding countries that may not otherwise be covered in existing surveillance efforts. Additionally, the Naval Health Research Center (NHRC) in San Diego, CA collects specimens from DoD recruit training centers and conducts surveillance along the Mexico border.

Landstuhl Regional Medical Center (LRMC) and Tripler Army Medical Center (TAMC) assist the program by processing DoD specimens for the EUCOM region and the State of Hawaii, respectively. This process seeks to provide more timely results and efficient transport of specimens.

Available on our website (listed below) is a list of previous weekly surveillance reports, program information (including an educational briefing and instruction pamphlets for clinic staff), and a dashboard containing respiratory data for our sentinel sites.

Errata:





#### Collaborating Partners

In addition to all participating DoD military sentinel sites, collaborating laboratories and medical centers (described above) may be further understood by reviewing the sites' website.

Click on the sites' icon to be directed to their webpage.













